78. Versatile Stereoselective Synthesis of Completely Protected Trifunctional α-Methylated α-Amino Acids Starting from Alanine

by Eva Altmann¹), Kurt Nebel¹), and Manfred Mutter*

Section de chimie de l'Université de Lausanne, 2, rue de la Barre, CH-1005 Lausanne

(17.IV.91)

A new route to completely protected α -methylated α -amino acids starting from alanine is described (see *Scheme*). These derivatives, which are obtained *via* base-catalyzed opening of the oxazolidinones (2*S*,4*R*)- and (2*R*,4*S*)-**2**, can be directly employed in peptide synthesis. The synthesis of both enantiomers of Z-protected α -methylaspartic acid β -(*tert*-butyl)ester (O^4 -(*tert*-butyl) hydrogen 2-methylaspartates (*R*) or (*S*)-**4a**), α -methyl-glutamic acid *y*-(*tert*-butyl) ester (O^5 -(*tert*-butyl) hydrogen 2-methylglutamate (*R*)- or (*S*)-**4b**), and of N^e -bis-Boc-protected α -methyllysine (N^6 , N^6 -bis[(*tert*-butyloxy)carbonyl]-2-methyllysine (*R*)- or (*S*)-**4c**) is described in full detail.

Introduction. – In recent years, the *de novo* design of peptides and proteins with predetermined secondary and tertiary structures has rapidly become a subject of major interest and importance in the area of bioorganic chemistry [1]. One of the chemical tools that are potentially available for structure stabilization in peptides is the incorporation of α -methylated α -amino acids [2]; due to severe restrictions of the rotational freedom around their N-C(α) and C(α)-C=O bond [3], α -methylated α -amino acids may be generally expected to display helix-inducing properties, as has been explicitly demonstrated for 2-aminoisobutyric acid (Aib) [4] and (S)-2-amino-2-methylbutyric acid = (S)-isovaline; (S)-Iva [2a, b]. However, although several routes for the stereoselective synthesis of these conformationally restricted amino acids have been reported over the last few years [5], we found that a convenient method for the direct synthesis of enantiomerically pure protected derivatives, which are crucial for the incorporation of these unusual building blocks into peptides, is still missing²). We have, therefore, now developed a simple but versatile synthetic procedure for the preparation of chiral α -methylated amino acids that are suitably protected for use in peptide synthesis.

Results and Discussion. – The Scheme shows our general strategy for the synthesis of the above compounds, which takes advantage of the 'principle of self reproduction of chirality centers' introduced by Seebach and coworkers [5b], a method displaying several advantageous features. The starting material in all our syntheses was either D- or L-alanine, depending on which enantiomer of the α -methylated amino acid was desired³).

¹) Present address: *Ciba-Geigy AG*, CH-4002 Basel.

²) We have reported previously the synthesis of H-(S)-Ser(2-Me-O-Bu')-OH starting from the α -methylated α -amino acid [6].

³) Only the synthesis starting from D-alanine is described in the *General Part*; for the analogous enantiomeric series starting from L-alanine, see *Exper. Part*.



a) Benzaldehyde, CH_2Cl_2 , reflux.

b) Benzyl chloroformate, 0° to r.t.

c) LHMDS or LDA, alkyl halide, -78° for 3 h, then to r.t. over night.

d) LDA; $CH_2 = CHCO_2Bu^t - 78^\circ$ for 3 h, then to r.t. over night.

e) 2 Equiv. of NaOH or LiOH, MeOH/H₂O 10:1, r.t., 30 min.

f) 2 Equiv. of NaOH or LiOH, MeOH/H₂O 1:1, 45°, 1 h.

g) H₂, Pd/C, MeOH, 1 h, r.t.

D-Alanine was first converted to the *Schiff* base with benzaldehyde followed by cyclization to the oxazolidinone (2S,4R)-1 by addition of benzyl chloroformate. Alkylations of (2S,4R)-1 were performed with lithium bis(trimethylsilyl)amide (LHMDS) or lithium diisopropylamide (LDA) as base and *tert*-butyl bromoacetate [7] and I(CH₂)₄N(Boc)₂ as electrophiles; they proceeded with excellent stereoselectivity⁴) with attack of the electrophile from the face opposite to the phenyl group (\rightarrow (2S,4R)-**2a** and (2S,4R)-**2c**, resp.). Oxazolidinone (2S,4R)-**2b**, the precursor for the α -methylglutamic acid γ -(*tert*-butyl) ester, was synthesized *via Michael* addition of *tert*-butyl acrylate to (2S,4R)-**1**. Product (2S,4R)-**2b** was formed in low yield (26%) but with high diastereoselectivity⁵). Addition of DMPU (N,N'-dimethylpropyleneurea) [8] suppressed the 1,4-addition almost quantitatively, an effect that had also been observed previously by *Seebach* and coworkers [9].

Oxazolidinone (2S,4R)-**2b** could be crystallized, and a computer-generated drawing of the X-ray structure is given in the *Figure*. Crystals of (2S,4R)-**2b** grown from Et₂O/pen-

⁴) Only one diastereoisomer could be detected by NMR spectroscopy of the purified oxazolidinones (2S,4R)-2a and (2S,4R)-2c and of their enantiomers. At a later stage, the diastereoisomeric purity of dipeptides Fmoc-Ala-X-OH (X = (S)-Asp(2-Me), (R)-Asp(2-Me), (S)-Lys(2-Me), (R)-Lys(2-Me)) was shown to be > 99% by HPLC. The synthesis of the peptides incorporating these α -methylated amino acids will be published elsewhere.

⁵) Only one diastereoisomer could be detected by NMR spectroscopy of the purified oxazolidinone (2S,4R)-2b and of its enantiomer. At a later stage, the diastereoisomeric purity of dipeptides Fmoc-Ala-X-OH (X = (S)-Glu(2-Me), (R)-Glu(2-Me)) was shown to be 98% by HPLC.



tane are triclinic, space group P1, containing two molecules (**A** and **B**) per asymmetric unit. The structure was solved routinely using direct methods⁶). The results of the X-ray structural analysis confirm our stereochemical assignments of oxazolidinones (2*S*,4*R*)-**2a**–c, which were originally inferred from the crystal structure of a related oxazolidinone

⁶) Crystal data for oxazolidinone (2S,4R)-2b: C₂₅H₂₉NO₆, triclinic, space group P1 with a = 13.743(2), b = 9.817(1), c = 9.906(1) Å, α = 112.0(1)°, β = 98.4(1), γ = 99.8(1)°; Z = 2, D_c = 1.24 gr ⋅ cm⁻³. On a Philips PW 1100 diffractometer, 5684 reflections were collected in the θ-2θ scan mode to 2θ = 56°, using graphite-monochromatized MoKα radiation (λ = 0.7107 Å). The structure was solved by direct methods using the SHELXS 86 program and refined by blocked least squares. The thermal parameters of all non-H-atoms were anisotropic. H-Atoms, partially found on a ΔF map and partially calculated were not refined. The final conventional R factor was 0.066 for the 1988 reflections considered observed [F > 7σ(F)]; R_w was 0.07 with w = 1/(σ²F + 0.0018F²). Refined atomic coordinates, anisotropic displacement parameters, bond lengths and angles have been deposited with the Cambridge Crystallographic Data Centre.

[5c] prepared by the same method. Further conformation came from the comparison of the optical rotation of (S)-2-methylaspartic acid (obtained from (S)-4a by hydrogenation and subsequent treatment with CF₃COOH) with literature data [10].

Key step in our synthesis is the base-promoted ring opening of the oxazolidinones (2S,4R)-2 leading to protected amino-acid derivatives (R)-3 and (R)-4³) which was carried out with NaOH [11] or LiOH in MeOH/H₂O mixtures. Depending on the reaction conditions, the Z-protected amino-acid esters (R)-3a-c or the Z-protected amino acids (R)-4a-c could be obtained selectively. Catalytic hydrogenation of (R)-3a gave access to the amino-acid ester (R)-5a in quantitative yield.

We are currently extending this new approach to protected derivatives of other trifunctional α -methylated α -amino acids. At the same time, we are synthesizing peptides containing different α -methylated α -amino acids in order to evaluate the putative β -turn and helix-stabilizing properties of these unusual building blocks.

Experimental Part

1. General. Reagents and solvents were purified by standard procedures [12]. All reactions involving Li derivatives were carried out under Ar. All chemicals (unless otherwise noted) were purchased from *Fluka AG*, Buchs, Switzerland. TLC: *Merck* precoated silica gel 60 *F-254* plates; detection with UV light (254 nm) if possible, and/or by development with 20% phosphomolybdic acid in EtOH and/or Cl₂/starch/KI. Flash chromatography (FC): silica gel 60 (230–400 mesh; 0.04–0.063 mm, *Merck*); according to [13]. HPLC analysis: *Waters* HPLC system; *Vydac C₁₈* column (25 × 0.4 cm) using H₂O (0.09% of CF₃COOH)/90% aq. MeCN (0.09% of CF₃COOH) as eluants. M.p.: uncorrected. [α]_D: *Perkin-Elmer-241* polarimeter. ¹H-NMR and ¹³C-NMR spectra: *Bruker-250-FT* (250 MHz) and *Bruker-WH-360-FT* (360 MHz) spectrometer; δ in ppm rel. to TMS, *J* in Hz. MS: *Nermag R* 10-10C (chemical ionisation (CI)) and *Finnigan-1020* (fast-atom bombardement (FAB)) mass spectrometer.

2. (2S,4R)-3-[(Benzyloxy)carbonyl]-4-methyl-2-phenyl-1,3-oxazolidin-5-one ((2S,4R)-1). To a suspension of 12.5 g (0.112 mol) of sodium D-alaninate in 500 ml of dry CH₂Cl₂, 11.31 ml (0.112 mol) of benzaldehyde were added, and the mixture was refluxed using a *Dean-Stark* apparatus for 21 h. It was then cooled to 0°, 14 ml (0.110 mol) of benzyl chloroformate were added, and stirring was continued at 0° for 5 h and then overnight at 25°. The solvent was evaporated, the resulting residue dissolved in 500 ml of AcOEt and successively washed with 5% NaHCO₃ soln., 5% KHSO₄ soln., and H₂O. The org. layer was dried (Na₂SO₄) and evaporated. The crude product (a yellow oil) was dried under high vacuum and analyzed by ¹H-NMR and HPLC: *cis/trans*-isomers 1:2.5. Al-though the *cis*- and *trans*-isomers were not separable by TLC, other impurities were efficiently removed by FC with CH₂Cl₂/pentane 1:1. Separation of the two isomers was subsequently achieved by crystallization from (i-Pr)₂O at -18° yielding 9.8 g (30.5%) of pure *trans*-oxazolidinone (2*S*,4*R*)-1. M.p. 77.3-78.4° [α]²⁵/₂₅ = -85.77 (*c* = 0.56, CH₂Cl₂). ¹H-NMR (C₂D₂Cl₄, 60°): 7.45-6.90 (*m*, 10 arom. H); 6.47 (*s*, H-C(2)); 5.10-4.92 (*m*, PhCH₂); 4.51 (*q*, *J* = 6.8, H-C(4)); 1.67 (*d*, *J* = 6.8, Me-C(4)). ¹³C-NMR (C₂D₂Cl₄, 60°): 172.07; 151.80; 136.47; 135.20; 129.96; 128.76; 128.35; 128.14; 127.74; 126.36; 89.29; 67.46; 51.95; 16.90. FAB-MS (C₁₈H₁₇NO₄ (311.34)): 312 ([*M* + 1]⁺).

3. Alkylations of Oxazolidinone Enolates with Alkyl Halides. General Procedure A. A soln. of 10 mmol of 1,1,1,3,3,3-hexamethyldisilazane (HMDS) in 40 ml of THF was cooled to -78° , and 6.25 ml of 1.6M BuLi were added dropwise. The resulting soln. was stirred for 10 min at -78° and then transferred (*via* cannula) to a precooled (-78°) soln. of 7.5 mmol of oxazolidinone (2*S*,4*R*)-1 in 40 ml of THF. The slightly yellow enolate soln. was stirred for 5–10 min at -78° , and then 9 mmol of alkyl halide were added. The mixture was stirred for 3 h at -78° and then 9 mmol of alkyl halide were added. The mixture was stirred for 3 h at -78° and then 9 mmol of alkyl halide were added. The mixture was stirred for 3 h at -78° and then allowed to warm to r.t. over night. THF was evaporated, the residue particult between sat. aq. NH₄Cl soln. and Et₂O, the aq. layer separated and extracted twice with Et₂O, and the combined ether extract dried (Na₂SO₄) and evaporated to give the crude product. General Procedure B. As described in General Procedure A, but with (i-Pr)₂NH instead of HMDS. The addition of DMPU [10] (10–12 ml) to the mixture in Procedure A or B had no effect on yields.

(2S,4R)-3-[(Benzyloxy)carbonyl]-4-{[(tert-butyloxy)carbonyl]methyl}-4-methyl-2-phenyl-1,3-oxazolidin-5-one ((2S,4R)-2a). From (2S,4R)-1 and tert-butyl bromoacetate according to Procedure A. FC (toluene/AcOEt 10:1→10:0.5) of the crude product yielded 76% of (2*S*,4*R*)-**2a**. Colourless oil. ¹H-NMR (C₂D₂Cl₄, 60°): 7.58–6.77 (*m*, 10 arom. H); 6.49 (*s*, H−C(2)); 5.21–4.89 (*m*, PhCH₂); 3.68 (*d*, J = 7, 1 H, CH₂−C(4)); 2.92 (*d*, J = 7, 1 H, CH₂−C(4)); 1.75 (*s*, Me−C(4)); 1.42 (*s*, *t*-Bu). ¹³C-NMR (C₂D₂Cl₄, 60°): 173.80; 168.93; 151.97; 136.96; 135.50; 129.72; 128.64; 128.47; 128.21; 127.83; 127.00; 89.81; 81.97; 67.48; 59.75; 41.65; 28.19; 28.06. FAB-MS (C₂A₄H₂₇NO₆ (425.48)): 426 ([*M* + 1]⁺).

(2S,4R)-3-[(Benzyloxy)carbonyl]-4-{4-{bis[(tert-butyloxy)carbonyl]amino}butyl}-4-methyl-2-phenyl-1,3-oxazolidin-5-one ((2S,4R)-2c). From (2S,4R)-1 and N,N-bis[(tert-butyloxy)carbonyl]-4-iodobutanamine (I(CH₂)₄N(Boc)₂) according to Procedure B. FC (100% toluene \rightarrow toluene/AcOEt 10:0.3) of the crude product yielded 76% of (2S,4R)-2c. Colourless oil. ¹H-NMR (C₂D₂Cl₄, 60°): 7.56-6.75 (m, 10 arom. H); 6.50 (s, H-C(2)); 5.30-4.82 (m, PhCH₂); 3.50 (t, CH₂N); 1.98-1.70 (m, CH₂); 1.76 (s, Me-C(4)); 1.65-1.40 (m, 2 CH₂); 1.52 (s, t-Bu); 1.30-1.20 (m, CH₂). FAB-MS (C₃₂H₄₃N₂O₈ (582.74)): 583 ([M + 1]⁺).

4. (2S,4R)-3-[(Benzyloxy)carbonyl]-4- $\{[(tert-butyloxy)carbonyl]ethyl]$ -4-methyl-2-phenyl-1,3-oxazolidin-5-one ((2S,4R)-**2b**). A soln. of 1.12 ml (8 mmol) of (i-Pr)₂NH in 40 ml of THF was cooled to -78° , and 5 ml of 1.6M BuLi in hexane were added dropwise. The resulting soln. was stirred for 10 min at -78° and then transferred *via* cannula to a precooled soln. of 2.35 g (7.5 mmol) of (2S,4R)-1. The dark yellow enolate soln. was stirred for 10 min at 78°, and then 1.2 ml (8 mmol) of *tert*-butyl acrylate were added, resulting in an immediate decolourisation of the enolate soln. The mixture was stirred for 3 h at -78° and then allowed to warm to r.t. overnight. THF was evaporated, the residue partitioned between sat. aq. NH₄Cl soln. and Et₂O, the aq. layer separated and twice extracted with Et₂O, and the combined org. extract dried (Na₂SO₄) and evaporated. FC (100% toluene \rightarrow toluene/ AcOEt 10:0.3) of the crude product yielded 0.880 g (26.5%) of (2S,4R)-**2b** as a colourless oil which was crystallized from Et₂O/pentane. M.p. 97.3–98.0°. [α]_D²⁵ = -52.49 (c = 0.65, CH₂Cl₂). ¹H-NMR (C₂D₂Cl₄, 60°): 7.56–6.76 (m, 10 arom. H); 6.51 (s, H–C(2)); 5.08 (m, PhCH₂); 2.62 (br. s, CH₂–C(4)); 2.20 (br. s, CH₂); 1.75 (s, Me–C(4)); 1.50 (s, t-Bu). ¹³C-NMR (C₂D₂Cl₄, 60°): 174.8; 172.5; 129.1; 128.2; 128.1; 81.2; 66.9; 59.9; 53.2; 32.2; 30.8; 28.5; 23.7. CI-MS (C₂₅H₂₉NO₆ (439.38)): 440 ([M + 1]⁺).

5. General Procedure C for the Preparation of N^{α} -(Benzyloxycarbonyl)- α -methyl-Substituted α -Amino Acid Methyl Esters (R)-3. To a soln. of oxazolidinone (2S,4R)-2 in 8–10 ml of MeOH were added 2 equiv. of 4N aq. NaOH (or LiOH), and the mixture was stirred at r.t. for 30 min. It was then diluted with 50 ml of H₂O, and the aq. layer was extracted 3 times with AcOEt. The combined org. extracts were dried (Na₂SO₄) and evaporated to give the crude product.

O⁴-(tert-*Butyl*) O¹-*Methyl* (R)-N²-*f*(*Benzyloxy*)*carbonyl*]-2-*methylaspartate* (Z-(R)-Asp(2-Me,O-Bu⁴)-OMe; (R)-3a) was obtained according to the *General Procedure* C from 3 g (7 mmol) of (2S,4R)-2a using LiOH. The crude product was purified by FC (toluene/AcOEt/AcOH 100:10:1): 2.45 g (81%) of (R)-3a, which was crystallized from pentane. M.p. 42.1-43.2°. $[\alpha l_{D5}^{25} = +12.04 (c = 0.30, CH_2Cl_2).$ ¹H-NMR (CDCl₃): 7.31 (*s*, 5 arom. H); 6.10 (br. *s*, NH); 5.08 (*s*, PhCH₂); 3.70 (*s*, CO₂CH₃); 3.21 (*d*, *J* = 7.0, 1 H, CH₂(3)); 2.92 (*d*, *J* = 7, 1 H, CH₂(3)); 1.60 (*s*, *t*-Bu). ¹³C-NMR (CDCl₃): 177.9; 169.6; 154.0; 136.1; 129.8; 128.3; 81.8; 61.4; 57.4; 42.1; 28.0; 23.1. CI-MS (C₁₈H₂₅NO₆ (337.18)): 338 ([*M* + 1]⁺).

 O^{5} -(tert-Butyl) O^{1} -Methyl (R)-N²-f(Benzyloxy)carbonyl]-2-methylglutamate (Z-(R)-Glu(2-Me,O-Bu¹)-OMe; (R)-3b) was obtained according to the General Procedure C from 4.39 g (10 mmol) of (2S,4R)-2b using LiOH. The crude product was purified by FC (toluene/AcOEt/AcOH 100:10:1): 2.84 g (78%) of (R)-3b. Colourless oil. $[\alpha]_{D}^{25} = +3.75$ (c = 0.80, MeOH). ¹H-NMR (CDCl₃): 7.36 (s, 5 arom. H); 5.78 (br. s, NH); 5.10 (s, PhCH₂); 3.75 (s, CO₂CH₃); 2.48–2.05 (m, 2 CH₂); 1.59 (s, Me–C(2)); 1.45 (s, t-Bu). ¹³C-NMR (CDCl₃): 174.5; 172.3; 128.9; 128.2; 81.1; 71.9; 60.0; 53.2; 32.1; 28.8; 23.7. CI-MS (C₁₉H₂₅NO₆ (365.42)): 366 ([M + 1]⁺).

Methyl (R)-N²-[(*Benzyloxy*)carbonyl]-N⁶,N⁶-bis[(tert-butyloxy)carbonyl]-2-methyllysine (Z-(R)-Lys(2-Me,N,N-Boc₂)-OMe; (R)-3c) was obtained according to the *General Procedure* C from 2.32 g (4 mmol) of (2S,4R)-2c using LiOH. The crude product was purified by FC (toluene/AcOEt/AcOH 100:10:1): 1.58 g (78%) of (R)-3c. Colourless oil. [α]₂₅²⁵ = +3.84 (c = 0.26, CH₂Cl₂). ¹H-NMR (CDCl₃): 7.29 (s, 5 arom. H); 5.65 (br. s, NH); 5.08 (s, PhCH₂); 3.70 (s, CO₂CH₃); 3.50 (t, CH₂N); 1.78 (m, 1 H, CH₂); 1.75 (m, 1 H, CH₂); 1.53 (s, Me–C(2)); 1.49 (s, 2 t-Bu); 1.52–1.45 (m, CH₂); 1.07–1.02 (m, CH₂). Cl-MS (C₂₆H₄₀NO₈ (508.57)): 509 ([M + 1]⁺).

6. General Procedure D for the Preparation of N^{α} -(Benzyloxycarbonyl)- α -methyl-Substituted α -Amino Acids (R)-4. To a soln. of 5 mmol of oxazolidinone (25,4R)-2 in 4 ml of MeOH were added 2 equiv. of 2N aq. NaOH (or LiOH); and the mixture was stirred for 1 h at 45°. It was then diluted with 50 ml of H₂O, and the aq. layer was extracted 3 times with Et₂O. The aq. layer was cooled to 0° and the pH adjusted to 3 with 2N HCl. The acidic aq. soln. was extracted 5 times with AcOEt, the org. layers were combined, dried (Na₂SO₄), and evaporated.

O⁴-(tert-*Butyl*) Hydrogen (R)-N²-[(Benzyloxy)carbonyl]-2-methylaspartate (Z-(R)-Asp(2-Me,O-Bu¹); (R)-4a) was obtained according to the General Procedure D from 3 g (7 mmol) of (2S,4R)-2a using LiOH. The crude product was purified by FC (toluene/AcOEt/AcOH 100:20:3): 1.80 g (76%) of (R)-4a, which was crystallized from CHCl₃/pentane. M.p. 88.2–89°. [α]_D = -3.84 (c = 0.26, CH₂Cl₂). ¹H-NMR (CDCl₃): 7.31 (s, 5 arom. H); 6.10 (br. s, NH); 5.09 (d, PhCH₂); 3.12 (d, J = 7.0, 1 H, CH₂(3)); 2.92 (d, J = 7, 1 H, CH₂(3)); 1.69 (s, Me–C(2)); 1.40 (s, t-Bu). ¹³C-NMR (CDCl₃): 177.9; 169.6; 154.0; 136.1; 129.9; 128.1; 128.0; 81.8; 61.9; 57.5; 42.1; 27.9; 23.2. CI-MS (C₁₂H₂₃NO₆ (337.35)): 338 ([M + 1]⁺).

O⁵-(tert-Butyl) Hydrogen (R)-N²-[(Benzyloxy)carbonyl]-2-methylglutamate (Z-(R)-Glu(2-Me,O-Bu'); (R)-4b) was obtained according to the General Procedure D from 3.10 g (7 mmol) of (2S,4R)-2b using NaOH. The crude product was purified by FC (toluene/AcOEt/AcOH 100:20:3): 1.9 g (77%) of (R)-4b. Colourless oil. $[\alpha]_{25}^{D5} = -1.15$ (c = 0.70, CH₂Cl₂). ¹H-NMR (CDCl₃): 7.35 (s, 5 arom. H); 5.98 (br. s, NH); 5.08 (s, PhCH₂); 2.42–2.07 (m, CH₂(3), CH₂(4)); 1.60 (s, Me–C(2)); 1.42 (s, t-Bu). CI-MS (C₁₈H₂₅NO₆(351.22)): 352 ([M + 1]⁺).

(R)-N²-*f* (Benzyloxy)carbonyl]-N⁶,N⁶-bisf (tert-butyloxy)carbonyl]-2-methyllysine (Z-(R)-Lys(2-Me,N,N, Boc₂); (R)-4c) was obtained according to the General Procedure D from 5.83 g (10 mmol) of (2S,4R)-2c using NaOH. The crude product was purified by FC (toluene/AcOEt/AcOH 100:20:3): 3.13 g (63.5%) of (R)-4c, which was crystallized from CHCl₃/pentane. M.p. 215–219° (dec.). $[\alpha]_{25}^{D5} = +7.12$ (c = 0.66, MeCN). ¹H-NMR (CDCl₃): 7.32 (s, 5 arom. H); 5.72 (br. s, NH); 5.10 (s, PhCH₂); 3.51 (t, CH₂N); 2.38–1.10 (m, 3 CH₂); 1.50 (s, 2 t-Bu). CI-MS (C₂₅H₃₈N₂O₈ (494.63)): 495 ([M + 1]⁺).

7. O⁴-(tert-*Butyl*) O¹-*Methyl* (R)-2-*Methylaspartate* ((R)-Asp(2-Me,O-Bu^t)-OMe; (R)-5a). To a soln. of 125 mg (0.37 mmol) of (R)-3a in 3 ml of MeOH, 22.5 mg of 10% Pd/C was added. The mixture was hydrogenated for 1 h, then the catalyst was filtered and the solvent evaporated. The product was briefly dried under high vacuum (caution, (R)-5a is extremely volatile): 76.5 mg (95%) of (R)-5a. ¹H-NMR (CDCl₃): 3.72 (s, CO₂CH₃); 2.90 (d, J = 6.0, 1 H, CH₂(3)); 2.02 (br. s, NH₂); 1.43 (s, t-Bu); 1.32 (s, Me-C(2)). CI-MS (C₁₀H₁₉NO₄ (217.12)): 218 ([M + 1]⁺).

8. N, N-Bis[(tert-butyloxy)carbonyl]-4-iodobutanamine (= Di(tert-butyl) [(4-Iodobutyl)imino]dicarboxylate; I(CH₂)₄N(Boc)₂). To 5 g of di(tert-butyl) iminodicarboxylate in 10 ml of EtOH were added 1.3 g (23 mmol) of KOH in 10 ml of EtOH. The mixture was stirred for 40 min at r.t., and the product was precipitated with dry Et₂O, collected by filtration, and dried under high vacuum. 5.75 g (98%) of di(tert-butyl) (potassioimino)dicarboxylate which was directly used for the next step without further purification. To a soln. of 5.75 g (22.5 mmol) of the K salt in 12 ml of DMF and 50 ml of dry CH₂Cl₂, 2.95 ml (25 mmol) of 1,4-dibromobutane were added, and the mixture was stirred for 3.5 h at 50°. After cooling to r.t., the mixture was filtered and the CH₂Cl₂ evaporated. The residue was taken up in 350 ml of AcOEt and washed with brine. The org. layer was dried (MgSO₄) and evaporated. Purification by FC (pentane/Et₂O 1:1) yielded 6.2 g (78.2%) of the bromide as a colourless oil. Conversion of the bromide to the iodide was carried out in 15 ml of dry acetone with 3.75 g (25 mmol) of NaI. The mixture was stirred for 24 h at r.t. in the dark, then the acetone was evaporated and the residue dissolved in 350 ml of Et₂O. The soln. was washed with H₂O, dried (MgSO₄), and evaporated. The residual oil was purified by short-column chromatography (Et₂O/penante 1:1): 6.2 g (88%) of the title compound, colourless oil.

Data of N, N-Bis[(tert-butyloxy)carbonyl]-4-bromobutanamine: ¹H-NMR (CDCl₃): 3.60 (t, CH₂N); 3.40 (t, CH₂Br); 1.88 (m, CH₂); 1.72 (m, CH₂); 1.50 (s, 2 t-Bu). CI-MS (C₁₄H₂₆NO₄Br (352.207)): 352 (M^+), 354 ($[M + 2]^+$).

Data of N,N-Bis[(tert-butyloxy)carbonyl]-4-iodobutanamine. ¹H-NMR (CDCl₃): 3.60 (t, CH₂); 3.18 (t, CH₂I); 1.80 (m, CH₂); 1.72 (m, CH₂); 1.50 (s, 2 t-Bu). CI-MS (C₁₄H₂₆NO₄I (399.203)): 400 ($[M + 1]^+$).

9. Enantiomeric Series. Experimental procedures and NMR data for (2R,4S)-1, (2R,4S)-2a-c, (S)-3a-c, (S)-4a-c, and (S)-5a are identical with those provided fro the corresponding enantiomeric compounds (see above).

(2R,4S)-1: M.p. 77.2-78.3°. $[\alpha]_{D}^{25} = +105.3$ (c = 0.45, CH₂Cl₂). (2R,4S)-**2b**: M.p. 97.4-98.0°. $[\alpha]_{D}^{25} = +52.96$ (c = 0.48, CH₂Cl₂). (S)-**3a**: M.p. 42.1-43.2°. $[\alpha]_{D}^{25} = -13.36$ (c = 0.37, CH₂Cl₂). (S)-**3b**: $[\alpha]_{D}^{25} = -8.36$ (c = 0.66, MeOH). (S)-**3c**: $[\alpha]_{D}^{25} = -4.63$ (c = 0.45, CH₂Cl₂). (S)-**4a**: M.p. 88.2-89.0°. $[\alpha]_{D}^{25} = +4.72$ (c = 0.46, CH₂Cl₂). (S)-**4b**: $[\alpha]_{D}^{25} = +1.14$ (c = 0.71, CH₂Cl₂). (S)-**4c**: M.p. 215-210° (dec.). $[\alpha]_{D}^{25} = -4.51$ (c = 0.27, CH₂Cl₂).

We are grateful for financial support by the Swiss National Science Foundation. The authors wish to thank Dr. G. Valle, Biopolymer Research Centre, C.N.R., Department of Organic Chemistry, University of Padova, Italy, for having solved the X-ray diffraction structure and Prof. C. Toniolo, University of Padova, Italy, for his collaboration.

REFERENCES

- a) M. Mutter, S. Vuilleumier, Angew. Chem. Int. Ed. 1989, 28, 535; b) W. F. DeGrado, Z. A. Wasserman, J. D. Lear, Science 1989, 243, 622; c) W. F. DeGrado, Adv. Prot.Chem. 1989, 39, 59; d) J. S. Richardson, D. C. Richardson, in 'Protein Engineering', Eds. D. L. Oxender and C. F. Fox, A. R. Liss, New York, 1987, p. 149; e) R. Moser, S. Frey, K. Münger, T. Helgans, S. Klauser, H. Langen, E. Winnacker, R. Merz, B. Gutte, Protein Eng. 1987, 1, 339.
- [2] a) E. Altmann, K.-H. Altmann, K. Nebel, M. Mutter, Int. J. Pept. Protein Res. 1988, 32, 344; b) G. Valle, M. Crisma, C. Toniolo, R. Beisswenger, A. Rieker, G. Jung, J. Am. Chem. Soc. 1989, 111, 6828; c) I. L. Karle, P. Balaram, Biochemistry 1990, 29, 6747; d) B. V. Venktataram Prasad, P. Balaram, CRC Crit. Rev. Biochem. 1984, 16, 307; e) C. Toniolo, E. Benedetti, ISI Atlas of Science Biochemistry 1988, 1, 225.
- [3] a) E. E. Hodgkin, J. D. Clark, K. R. Miller, G. R. Marshall, *Biopolymers* **1990**, *30*, 533; b) A. W. Burgess, S. J. Leach, *ibid*. **1973**, *12*, 2599; c) Y. Paterson, S. M. Rumsey, E. Benedetti, G. Némethy, H. A. Scheraga, J. Am. Chem. Soc. **1981**, *103*, 2947; d) V. Barone, F. Lelj, A. Baroso, B. DiBlasio, P. Grimaldi, V. Pavone, C. Pedone, Biopolymers **1985**, *24*, 1759.
- [4] a) R. Bosch, G. Jung, H. Schmitt, W. Winter, *Biopolymers* 1985, 24, 961; b) V. Pavone, B.A. DiBlasio, A. Santini, E. Benedetti, C. Pedone, C. Toniolo, M. Crisma, J. Mol. Biol. 1990, 214, 633.
- [5] a) D. Seebach, E. Dziadulewicz, L. Behrendt, S. Cantoreggi, R. Fitzi, *Liebigs Ann. Chem.* 1989, 1215; b) D. Seebach, R. Imwinkelried, T. Weber, in 'Modern Synthetic Methods', Ed. R. Scheffold, Springer Verlag, Heidelberg, 1986, Vol. 4, p. 125; c) K. Nebel, M. Mutter, *Tetrahedron* 1988, 44, 4793; d) U. Schöllkopf, in 'Topics in Current Chemistry', Ed. F.L. Boschke, Springer Verlag, Heidelberg, 1983, Vol. 109, p. 65; e) K. Weinges, H. Brachmann, P. Stahnecker, H. Rodewald, M. Nixdorf, H. Irmgartinger, *Liebigs Ann. Chem.* 1985, 566; f) I. Ojima, C.C. Hauh-Jyun, Q. Xianogang, *Tetrahedron* 1988, 44, 5307; g) R.M. Williams, in 'Synthesis of Optically Active α-Amino Acids', Eds. J. E. Baldwin and P.D. Magnus, Pergamon Press, Oxford, 1989.
- [6] E. Altmann, K.-H. Altmann, M. Mutter, Angew. Chem. Int. Ed. 1988, 27, 858.
- [7] A. Fadel, J. Salaün, Tetrahedron Lett. 1987, 28, 2243.
- [8] a) D. Seebach, R. Henning, T. Mukhopadhyay, Chem. Ber. 1982, 115, 1765; b) T. Mukhopadhyay,
 D. Seebach, Helv. Chim. Acta 1982, 65, 3851.
- [9] D. Seebach, M. Boes, R. Naef, W. B. Schweizer, J. Am. Chem. Soc. 1983, 105, 5390.
- [10] J.D. Aebi, D. Seebach, Helv. Chim. Acta 1985, 68, 1507.
- [11] S. Karady, J.S. Amato, L.M. Weinstock, Tetrahedron Lett. 1984, 25, 4337.
- [12] D. D. Perrin, W. L. Armarego, in 'Purification of Laboratory Chemicals', 3rd edn., Pergamon Press, Oxford, 1988.
- [13] W.C. Still, M. Kahn, A. Mitra, J. Org. Chem. 1978, 43, 2923.